

# Package ‘ModTools’

July 31, 2024

**Type** Package

**Title** Tools for Building Regression and Classification Models

**Version** 0.9.12

**Date** 2024-07-31

**Encoding** UTF-8

**Description**

Collection of tools for regression and classification tasks. The package implements a consistent user interface to the most popular regression and classification algorithms, such as random forest, neural networks, C5 trees and support vector machines, and complements it with a handful of auxiliary functions, such as variable importance and a tuning function for the parameters.

**Depends** DescTools, MASS, nnet, survival, R (>= 3.5.0)

**License** GPL (>= 2)

**Suggests** VGAM

**Imports** e1071, C50, rpart, randomForest, pROC, methods, relaimpo,  
rpart.plot, lattice, lmtest, car, robustbase, class,  
NeuralNetTools, naivebayes, sandwich, AER, boot

**URL** <https://andrisignorell.github.io/ModTools/>,  
<https://github.com/AndriSignorell/ModTools/>

**BugReports** <https://github.com/AndriSignorell/ModTools/issues>

**LazyLoad** yes

**LazyData** yes

**RoxygenNote** 7.3.2

**Config/testthat/edition** 3

**NeedsCompilation** no

**Author** Andri Signorell [aut, cre],  
Bernhard Compton [ctb],  
Marcel Dettling [ctb],  
Alexandre Hainard [ctb],  
Max Kuhn [ctb],  
Frédérique Lisacek [ctb],

Michal Majka [ctb],  
 Markus Müller [ctb],  
 Dan Putler [ctb],  
 Jean-Charles Sanchez [ctb],  
 Natalia Tiberti [ctb],  
 Natacha Turck [ctb],  
 Jarek Tuszynski [ctb],  
 Robin Xavier [ctb],  
 Achim Zeileis [ctb]

**Maintainer** Andri Signorell <andri@signorell.net>

**Repository** CRAN

**Date/Publication** 2024-07-31 05:50:10 UTC

## Contents

ModTools-package . . . . .	3
BestCut . . . . .	8
bioChemists . . . . .	9
BreuschPaganTest . . . . .	10
CoeffDiffCI . . . . .	11
CP . . . . .	12
d.glass . . . . .	14
d.pima . . . . .	15
FitMod . . . . .	16
LeafRates . . . . .	17
LogitBoost . . . . .	19
Node . . . . .	21
Over-/Undersample . . . . .	22
PlotLift . . . . .	23
predict.zeroinfl . . . . .	24
PredictCI . . . . .	26
RefLevel . . . . .	27
Response . . . . .	28
RobSummary . . . . .	29
ROC . . . . .	30
Rules . . . . .	31
SplitTrainTest . . . . .	32
TModC . . . . .	33
Tobit . . . . .	34
Tune . . . . .	35
VarImp . . . . .	37
zeroinfl . . . . .	38
zeroinfl.control . . . . .	42

**Index** **44**

## Description

There is a rich selection of R packages implementing algorithms for classification and regression tasks out there. The authors legitimately take the liberty to tailor the function interfaces according to their own taste and needs. For us other users, however, this often results in struggling with user interfaces, some of which are rather weird - to put it mildly - and almost always different in terms of arguments and result structures. **ModTools** pursues the goal of offering uniform handling for the most important regression and classification models in applied data analyses.

The function `FitMod()` is designed as a simple and consistent interface to these original functions while maintaining the flexibility to pass on all possible arguments. `print`, `plot`, `summary` and `predict` operations can so be carried out following the same logic. The results will again be reshaped to a reasonable standard.

For all the functions of this package Google styleguides are used as naming rules (in absence of convincing alternatives). The 'BigCamelCase' style has been consequently applied to functions borrowed from contributed R packages as well.

As always: Feedback, feature requests, bugreports and other suggestions are welcome!

## Details

The `ModTools::FitMod()` function comprises interfaces to the following models:

### Regression:

<code>lm()</code>	Linear model OLS ( <b>base</b> )
<code>lmrob()</code>	Robust linear model ( <b>robustbase</b> )
<code>poisson()</code>	GLM model with family poisson ( <b>base</b> )
<code>negbin()</code>	GLM model with family negative.binomial ( <b>MASS</b> )
<code>gamma()</code>	GLM model with family gamma ( <b>base</b> )
<code>tobit()</code>	Tobit model for censored responses (package <b>AER</b> )

### Classification:

<code>lda()</code>	Linear discriminant analysis ( <b>MASS</b> )
<code>qda()</code>	Quadratic discriminant analysis ( <b>MASS</b> )
<code>logit()</code>	Logistic Regression model <code>glm</code> , family binomial(logit)( <b>base</b> )
<code>multinom()</code>	Multinomial Regression model ( <b>nnet</b> )
<code>polr()</code>	Proportional odds model ( <b>MASS</b> )
<code>rpart()</code>	Regression and classification trees ( <b>rpart</b> )
<code>nnet()</code>	Neuronal networks ( <b>nnet</b> )
<code>randomForest()</code>	Random forests ( <b>randomForest</b> )
<code>C5.0()</code>	C5.0 tree ( <b>C50</b> )
<code>svm()</code>	Support vector machines ( <b>e1071</b> )
<code>naive_bayes()</code>	Naive Bayes classifier ( <b>naivebayes</b> )

`LogitBoost()` Logit boost (using decision stumps as weak learners) (**ModTools**)

**Preprocess:**

`SplitTrainTest()` Splits a data frame or index vector into a training and a test sample  
`OverSample()` Get balanced datasets by sampling with replacement.

**Manipulating rpart objects:**

`CP()` Extract and plot complexity table of an rpart tree.  
`Node()` Accessor to the most important properties of a node, being a split or a leaf.  
`Rules()` Extract the decision rules from top to the end node of an rpart tree.  
`LeafRates()` Returns the misclassification rates in all end nodes.

**Prediction and Validation:**

`Response()` Extract the response variable of any model.  
`predict()` Consistent predict for `FitMod` models  
`VarImp()` Variable importance for most `FitMod` models  
`ROC()` ROC curves for all dichotomous classification `FitMod` models  
`BestCut()` Find the optimal cut for a classification based on the ROC curve.  
`PlotLift()` Produces a lift chart for a binary classification model  
`TModC()` Aggregated results for multiple `FitMod` classification models  
`Tune()` Tuning approaches to find optimal parameters for `FitMod` classification models.  
`RobSummary()` Robust summary for GLM models (poisson).

**Tests:**

`BreuschPaganTest()` Breusch-Pagan test against heteroskedasticity.

**Warning**

This package is still under development. You should be aware that everything in the package might be subject to change. Backward compatibility is not yet guaranteed. Functions may be deleted or renamed and new syntax may be inconsistent with earlier versions. By release of version 1.0 the "deprecated-defunct process" will be installed.

**Author(s)**

Andri Signorell  
Helsana Versicherungen AG, Health Sciences, Zurich  
HWZ University of Applied Sciences in Business Administration Zurich.

Includes R source code and/or documentation previously published by (in alphabetical order):  
Bernhard Compton, Marcel Dettling, Max Kuhn, Michal Majka, Dan Putler, Jarek Tuszynski, Robin Xavier, Achim Zeileis

The good things come from all these guys, any problems are likely due to my tweaking. Thank you all!

Maintainer: Andri Signorell <andri@signorell.net>

## Examples

```
r.swiss <- FitMod(Fertility ~ ., swiss, fitfn="lm")
r.swiss
# PlotTA(r.swiss)
# PlotQQNorm(r.swiss)

## Count models

data(housing, package="MASS")

# poisson count
r.pois <- FitMod(Freq ~ Infl*Type*Cont + Sat, family=poisson, data=housing, fitfn="poisson")

# negative binomial count
r.nb <- FitMod(Freq ~ Infl*Type*Cont + Sat, data=housing, fitfn="negbin")
summary(r.nb)

r.log <- FitMod(log(Freq) ~ Infl*Type*Cont + Sat, data=housing, fitfn="lm")
summary(r.log)

r.ols <- FitMod(Freq ~ Infl*Type*Cont + Sat, data=housing, fitfn="lm")
summary(r.ols)

r.gam <- FitMod(Freq ~ Infl*Type*Cont + Sat, data=housing, fitfn="gamma")
summary(r.gam)

r.gami <- FitMod(Freq ~ Infl*Type*Cont + Sat, data=housing, fitfn="gamma", link="identity")
summary(r.gami)

old <-options(digits=3)
TMod(r.pois, r.nb, r.log, r.ols, r.gam, r.gami)
options(old)

## Ordered Regression

r.polr <- FitMod(Sat ~ Infl + Type + Cont, data=housing, fitfn="polr", weights = Freq)

# multinomial Regression
# r.mult <- FitMod(factor(Sat, ordered=FALSE) ~ Infl + Type + Cont, data=housing,
# weights = housing$Freq, fitfn="multinom")
```

```

# Regression tree
r.rp <- FitMod(factor(Sat, ordered=FALSE) ~ Infl + Type + Cont, data=housing,
               weights = housing$Freq, fitfn="rpart")

# compare predictions
d.p <- expand.grid(Infl=levels(housing$Infl), Type=levels(housing$Type), Cont=levels(housing$Cont))
d.p$polr <- predict(r.polr, newdata=d.p)
# ??
# d.p$ols <- factor(round(predict(r.ols, newdata=d.p)^2), labels=levels(housing$Sat))
# d.p$mult <- predict(r.mult, newdata=d.p)
d.p$rp <- predict(r.rp, newdata=d.p, type="class")

d.p

# Classification with 2 classes *****

r.pima <- FitMod(diabetes ~ ., d.pima, fitfn="logit")
r.pima
Conf(r.pima)
plot(ROC(r.pima))
OddsRatio(r.pima)

# rpart tree
rp.pima <- FitMod(diabetes ~ ., d.pima, fitfn="rpart")
rp.pima
Conf(rp.pima)
lines(ROC(rp.pima), col=hblue)
# to be improved
plot(rp.pima, col=SetAlpha(c("blue","red"), 0.4), cex=0.7)

# Random Forest
rf.pima <- FitMod(diabetes ~ ., d.pima, method="class", fitfn="randomForest")
rf.pima
Conf(rf.pima)
lines(ROC(r.pima), col=hred)

# more models to compare

d.pim <- SplitTrainTest(d.pima, p = 0.2)
mdiab <- formula(diabetes ~ pregnant + glucose + pressure + triceps
                + insulin + mass + pedigree + age)

r.glm <- FitMod(mdiab, data=d.pim$train, fitfn="logit")
r.rp <- FitMod(mdiab, data=d.pim$train, fitfn="rpart")
r.rf <- FitMod(mdiab, data=d.pim$train, fitfn="randomForest")
r.svm <- FitMod(mdiab, data=d.pim$train, fitfn="svm")
r.c5 <- FitMod(mdiab, data=d.pim$train, fitfn="C5.0")

```

```

r.nn <- FitMod(mdiab, data=d.pim$train, fitfn="nnet")
r.nb <- FitMod(mdiab, data=d.pim$train, fitfn="naive_bayes")
r.lda <- FitMod(mdiab, data=d.pim$train, fitfn="lda")
r.qda <- FitMod(mdiab, data=d.pim$train, fitfn="qda")
r.lb <- FitMod(mdiab, data=d.pim$train, fitfn="lb")

mods <- list(glm=r.glm, rp=r.rp, rf=r.rf, svm=r.svm, c5=r.c5
            , nn=r.nn, nb=r.nb, lda=r.lda, qda=r.qda, lb=r.lb)

# insight in the Regression tree
plot(r.rp, box.palette = as.list(Pal("Helsana", alpha = 0.5)))

# Insample accuracy ...
TModC(mods, ord="auc")
# ... is substantially different from the out-of-bag:
TModC(mods, newdata=d.pim$test, reference=d.pim$test$diabetes, ord="bs")
# C5 and SVM turn out to be show-offs! They overfit quite ordinary
# whereas randomforest and logit keep their promises. ...

sapply(mods, function(z) VarImp(z))

# Multinomial classification problem with n classes *****

d.gl <- SplitTrainTest(d.glass, p = 0.2)
mglass <- formula(Type ~ RI + Na + Mg + Al + Si + K + Ca + Ba + Fe)

# *** raises an unclear error in CRAN-Debian tests *** ??
# r.mult <- FitMod(mglass, data=d.gl$train, maxit=600, fitfn="multinom")
r.rp <- FitMod(mglass, data=d.gl$train, fitfn="rpart")
r.rf <- FitMod(mglass, data=d.gl$train, fitfn="randomForest")
r.svm <- FitMod(mglass, data=d.gl$train, fitfn="svm")
r.c5 <- FitMod(mglass, data=d.gl$train, fitfn="C5.0")
r.nn <- FitMod(mglass, data=d.gl$train, fitfn="nnet")
r.nbay <- FitMod(mglass, data=d.gl$train, fitfn="naive_bayes")
r.lda <- FitMod(mglass, data=d.gl$train, fitfn="lda")
# r.qda <- FitMod(mglass, data=d.glass, fitfn="qda")
r.lb <- FitMod(mglass, data=d.gl$train, fitfn="lb")

mods <- list(rp=r.rp, rf=r.rf, svm=r.svm, c5=r.c5,
            nn=r.nn, nbay=r.nbay, lda=r.lda, lb=r.lb)

# confusion matrix and other quality measures can be calculated with Conf()
Conf(r.rf)

# we only extract the general accuracy
sapply(lapply(mods, function(z) Conf(z)), "[[", "acc")

# let's compare r.mult with a model without RI as predictor
# Conf(r.mult)
# Conf(update(r.mult, . ~ . -RI))

```

---

**BestCut***Best Cutpoint for a ROC Curve*

---

**Description**

Returns the best cutpoint for a given classification model.

**Usage**

```
BestCut(x, method = c("youden", "closest.topleft"))
```

**Arguments**

x	a roc object from the roc function
method	one of "youden" or "closest.topleft", controls how the optimal threshold is determined. See details.

**Details**

The method argument controls how the optimal threshold is determined.

'youden' Youden's J statistic (Youden, 1950) is employed. The optimal cut-off is the threshold that maximizes the distance to the identity (diagonal) line. Can be shortened to "y".

The optimality criterion is:

$$\max(\text{sensitivities} + \text{specificities})$$

'closest.topleft' The optimal threshold is the point closest to the top-left part of the plot with perfect sensitivity or specificity. Can be shortened to "c" or "t".

The optimality criterion is:

$$\min((1 - \text{sensitivities})^2 + (1 - \text{specificities})^2)$$

**Value**

the threshold value

**Author(s)**

Robin Xavier <proC-cran@xavier.robin.name>, Andri Signorell <andri@signorell.net> (interface)

**References**

Xavier Robin, Natacha Turck, Alexandre Hainard, *et al.* (2011) "pROC: an open-source package for R and S+ to analyze and compare ROC curves". *BMC Bioinformatics*, **7**, 77. doi:10.1186/147121051277.



**See Also**[ROC](#)**Examples**

```
r.glm <- FitMod(diabetes ~ ., data = d.pima, fitfn="logit")  
  
ROC(r.glm)  
BestCut(ROC(r.glm))
```

---

bioChemists	<i>article production by graduate students in biochemistry Ph.D. programs</i>
-------------	---

---

**Description**

A sample of 915 biochemistry graduate students.

**Usage**

```
data(bioChemists)
```

**Format**

art count of articles produced during last 3 years of Ph.D.  
fem factor indicating gender of student, with levels Men and Women  
mar factor indicating marital status of student, with levels Single and Married  
kid5 number of children aged 5 or younger  
phd prestige of Ph.D. department  
ment count of articles produced by Ph.D. mentor during last 3 years

**References**

Long, J. Scott. 1990. The origins of sex differences in science. *Social Forces*. 68(3):1297-1316.  
Long, J. Scott. 1997. *Regression Models for Categorical and Limited Dependent Variables*. Thousand Oaks, California: Sage.

---

BreuschPaganTest      *Breusch-Pagan Test*

---

### Description

Performs the Breusch-Pagan test against heteroskedasticity.

### Usage

```
BreuschPaganTest(formula, varformula = NULL, studentize = TRUE, data = list())
```

### Arguments

formula	a symbolic description for the model to be tested (or a fitted "lm" object).
varformula	a formula describing only the potential explanatory variables for the variance (no dependent variable needed). By default the same explanatory variables are taken as in the main regression model.
studentize	logical. If set to TRUE Koenker's studentized version of the test statistic will be used.
data	an optional data frame containing the variables in the model. By default the variables are taken from the environment which BreuschPaganTest is called from.

### Details

The Breusch-Pagan test fits a linear regression model to the residuals of a linear regression model (by default the same explanatory variables are taken as in the main regression model) and rejects if too much of the variance is explained by the additional explanatory variables.

Under  $H_0$  the test statistic of the Breusch-Pagan test follows a chi-squared distribution with parameter (the number of regressors without the constant in the model) degrees of freedom.

Examples can not only be found on this page, but also on the help pages of the data sets [bondyield](#), [currencysubstitution](#), [growthofmoney](#), [moneydemand](#), [unemployment](#), [wages](#).

### Value

A list with class "htest" containing the following components:

statistic	the value of the test statistic.
p.value	the p-value of the test.
parameter	degrees of freedom.
method	a character string indicating what type of test was performed.
data.name	a character string giving the name(s) of the data.

### Author(s)

Achim Zeileis <Achim.Zeileis@R-project.org>

## References

- T.S. Breusch & A.R. Pagan (1979), A Simple Test for Heteroscedasticity and Random Coefficient Variation. *Econometrica* **47**, 1287–1294
- R. Koenker (1981), A Note on Studentizing a Test for Heteroscedasticity. *Journal of Econometrics* **17**, 107–112.
- W. Kraemer & H. Sonnberger (1986), *The Linear Regression Model under Test*. Heidelberg: Physica

## See Also

[lm](#), [ncvTest](#)

## Examples

```
## generate a regressor
x <- rep(c(-1,1), 50)

## generate heteroskedastic and homoskedastic disturbances
err1 <- rnorm(100, sd=rep(c(1,2), 50))
err2 <- rnorm(100)

## generate a linear relationship
y1 <- 1 + x + err1
y2 <- 1 + x + err2

## perform Breusch-Pagan test
BreuschPaganTest(y1 ~ x)
BreuschPaganTest(y2 ~ x)
```

---

CoeffDiffCI	<i>Confidence Interval for the Difference of Two Coefficients in a Linear Model</i>
-------------	---

---

## Description

Calculate the confidence interval for the difference of two coefficients in a linear model.

## Usage

```
CoeffDiffCI(x, coeff, conf.level = 0.95, sides = c("two.sided", "left", "right"))
```

## Arguments

x	the linear model object
coeff	a vector of length two, containing either the names or the index of the two coefficients whose difference should be used
conf.level	confidence level of the interval.

`sides` a character string specifying the side of the confidence interval, must be one of "two.sided" (default), "left" or "right". You can specify just the initial letter. "left" would be analogue to a hypothesis of "greater" in a t.test.

### Details

This is quite useful in the course of the modelling process.

### Value

a numeric vector with 3 elements:

<code>mean</code>	mean
<code>lwr.ci</code>	lower bound of the confidence interval
<code>upr.ci</code>	upper bound of the confidence interval

### Author(s)

Andri Signorell <andri@signorell.net>

### See Also

[linearHypothesis\(\)](#)

### Examples

```
# get some model first...
r.lm <- FitMod(Fertility ~ ., data=swiss, fitfn="lm")

# calculate the confidence interval for the difference of the
# coefficients Examination and Education
CoeffDiffCI(r.lm, c("Examination", "Education"))

# the test could be calculated as
car::linearHypothesis(r.lm, "Education = Examination")
```

### Description

Extracts, prints and plots the complexity table of an rpart model.

**Usage**

```
CP(x, ...)

## S3 method for class 'CP'
print(x, digits = getOption("digits") - 2L, ...)
## S3 method for class 'CP'
plot(x, minline = TRUE, lty = 3, col = 1,
      upper = c("size", "splits", "none"), ...)
```

**Arguments**

x	fitted model object of class "rpart". This is assumed to be the result of some function that produces an object with the same named components as that returned by the rpart function.
digits	the number of digits of numbers to print.
minline	whether a horizontal line is drawn 1SE above the minimum of the curve.
lty	line type for this line
col	colour for this line
upper	what is plotted on the top axis: the size of the tree (the number of leaves) ("size"), the number of splits ("splits") or nothing ("none").
...	further arguments passed to print and plot

**Details**

The complexity parameter table is hidden deep in the entrails of the rpart result object, it is convenient to have a function to extract it.

**Value**

A list containing the following components:

cp	the complexity table
x	the rpart object

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[printcp](#), [plotcp](#)

**Examples**

```
r.rp <- FitMod(diabetes ~ ., d.pima, fitfn="rpart")

CP(r.rp)
plot(CP(r.rp))
```

---

d.glass

*Measurements of Forensic Glass Fragments*

---

### Description

The d.glass data frame has 214 rows and 10 columns. It was collected by B. German on fragments of glass collected in forensic work.

### Usage

d.glass

### Format

This data frame contains the following columns:

RI refractive index; more precisely the refractive index is 1.518xxxx.

The next 8 measurements are percentages by weight of oxides.

Na sodium.

Mg manganese.

Al aluminium.

Si silicon.

K potassium.

Ca calcium.

Ba barium.

Fe iron.

Type The fragments were originally classed into seven types, one of which was absent in this dataset. The categories which occur are window float glass (WinF: 70), window non-float glass (WinNF: 76), vehicle window glass (Veh: 17), containers (Con: 13), tableware (Tabl: 9) and vehicle headlamps (Head: 29).

### References

Venables, W. N. and Ripley, B. D. (2002) *Modern Applied Statistics with S*. Fourth edition. Springer.

---

d.pima

*Diabetes survey on Pima Indians*

---

### Description

The National Institute of Diabetes and Digestive and Kidney Diseases conducted a study on 768 adult female Pima Indians living near Phoenix.

### Usage

```
data(d.pima)
data(d.pima2)
```

### Format

The dataset contains the following variables

pregnant Number of times pregnant

glucose Plasma glucose concentration at 2 hours in an oral glucose tolerance test

diastolic Diastolic blood pressure (mm Hg)

triceps Triceps skin fold thickness (mm)

insulin 2-Hour serum insulin (mu U/ml)

bmi Body mass index (weight in kg/(height in metres squared))

diabetes Diabetes pedigree function

age Age (years)

test test whether the patient shows signs of diabetes (coded 0 if negative, 1 if positive)

### Details

d.pima2 is the same dataset as d.pima with the only change, that invalid 0-values are replaced by NAs.

### Note

This dataset has been borrowed from Julian Faraway's package:  
*faraway*: Functions and datasets for books by Julian Faraway, 2015

### Source

The data may be obtained from the package MASS.

**Description**

Popular implementations of algorithms are characterized by partly unconventional implementations of the operating standards in R. For example, the function `e1071::SVM()` returns the predicted values as attributes!

`FitMod()` is designed as a wrapping function to offer a consistent interface for a selection of most often used classification and regression models.

**Usage**

```
FitMod(formula, data, ..., subset, na.action = na.pass, fitfn = NULL)

## S3 method for class 'FitMod'
predict(object, ...)
## S3 method for class 'FitMod'
plot(x, ...)
## S3 method for class 'FitMod'
summary(object, ...)
## S3 method for class 'FitMod'
drop1(object, ...)
```

**Arguments**

<code>x</code>	a fitted object of class "FitMod".
<code>formula</code>	a formula expression as for classification and regression models, of the form <code>response ~ predictors</code> . The response should be a factor or a matrix with <code>K</code> columns, which will be interpreted as counts for each of <code>K</code> classes. See the documentation of <code>formula()</code> for other details.
<code>data</code>	an optional data frame in which to interpret the variables occurring in formula.
<code>subset</code>	expression saying which subset of the rows of the data should be used in the fit. All observations are included by default.
<code>na.action</code>	a function to filter missing data.
<code>fitfn</code>	code for the fitting function to be used for regression or classifying. So far implemented are: <code>lm</code> , <code>lmrob</code> , <code>poisson</code> , <code>quasipoisson</code> , <code>gamma</code> , <code>negbin</code> , <code>poisson</code> , <code>polr</code> , <code>tobit</code> , <code>zeroinfl</code> , <code>multinom</code> , <code>poisson</code> , <code>rpart</code> , <code>randomForest</code> , <code>logit</code> , <code>nnet</code> , <code>C5.0</code> , <code>lda</code> , <code>qda</code> , <code>svm</code> , <code>naive_bayes</code> , <code>lb</code> .
<code>object</code>	the model object.
<code>...</code>	further arguments passed to the underlying functions.



**Details**

The function will in general return the original object, extended by a further class `FitMod`, which allows to capture the output and plot routines.

The classifying algorithms will at the minimum offer the predicting options `type = c("class", "prob")` additionally to those implemented by the underlying function.

**Value**

model object as returned by the calculating function extended with the `FitMod` class.

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[lm](#), [rpart](#)

**Examples**

```
r.lm <- FitMod(Fertility ~ ., data=swiss, fitfn="lm")

r.logit <- FitMod(diabetes ~ glucose + pressure + mass + age,
                 data=d.pima, fitfn="logit")

r.svm <- FitMod(diabetes ~ glucose + pressure + mass + age,
               data=d.pima, fitfn="svm")
```

---

LeafRates

*Leafrates for the Nodes of an 'rpart' Tree*

---

**Description**

Return the frequencies of correct and wrong classifications in given node(s) in tabular form. The 'purity', denoting the relative frequency of correctly classified elements, is a useful information for the interpretation of regression and classification trees and a measure for its quality.

**Usage**

```
LeafRates(x)

## S3 method for class 'LeafRates'
plot(x, col = NULL, which = c("rel", "abs"),
     layout = NULL, ylim = NULL, ...)
```

**Arguments**

x	fitted model object of class rpart.
col	color for the bars in the plot
which	one out of "rel" or "abs", denoting whether relative or absolute frequencies should be used for the plot.
layout	vector defining the layout
ylim	the y limits of the plot.
...	further arguments (not used).

**Details**

The result comprises absolute and relative frequencies per leaf.

**Value**

A list with 5 elements consisting of:

node	the node id (of the leaf)
freq	the absolute frequency of correct and wrong classifications
p.row	the relative frequency of correct and wrong classifications
mfreq	the total number of cases
mperc	the percentage of the sample in the leaf

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[Node, Rules](#)

**Examples**

```
r.rp <- FitMod(Species ~ ., data=iris, fitfn="rpart")
LeafRates(r.rp)

plot(LeafRates(r.rp))
```

---

 LogitBoost

*LogitBoost Classification Algorithm*


---

### Description

Train logitboost classification algorithm using decision stumps (one node decision trees) as weak learners.

### Usage

```
LogitBoost(x, ...)

## S3 method for class 'formula'
LogitBoost(formula, data, ..., subset, na.action)

## Default S3 method:
LogitBoost(x, y, nIter=ncol(x), ...)
```

### Arguments

formula	a formula expression as for regression models, of the form response ~ predictors. The response should be a factor or a matrix with K columns, which will be interpreted as counts for each of K classes. See the documentation of <a href="#">formula()</a> for other details.
data	an optional data frame in which to interpret the variables occurring in formula.
...	additional arguments for nnet
subset	expression saying which subset of the rows of the data should be used in the fit. All observations are included by default.
na.action	a function to filter missing data.
x	A matrix or data frame with training data. Rows contain samples and columns contain features
y	Class labels for the training data samples. A response vector with one label for each row/component of xlearn. Can be either a factor, string or a numeric vector.
nIter	An integer, describing the number of iterations for which boosting should be run, or number of decision stumps that will be used.

### Details

The function was adapted from logitboost.R function written by Marcel Dettling. See references and "See Also" section. The code was modified in order to make it much faster for very large data sets. The speed-up was achieved by implementing a internal version of decision stump classifier instead of using calls to [rpart](#). That way, some of the most time consuming operations were precomputed once, instead of performing them at each iteration. Another difference is that training and testing phases of the classification process were split into separate functions.

**Value**

An object of class "LogitBoost" including components:

Stump	List of decision stumps (one node decision trees) used: <ul style="list-style-type: none"> <li>• column 1: feature numbers or each stump, or which column each stump operates on</li> <li>• column 2: threshold to be used for that column</li> <li>• column 3: bigger/smaller info: 1 means that if values in the column are above threshold than corresponding samples will be labeled as lablist[1]. Value "-1" means the opposite.</li> </ul> <p>If there are more than two classes, than several "Stumps" will be cbind'ed</p>
lablist	names of each class

**Author(s)**

Jarek Tuszynski (SAIC) <jaroslav.w.tuszynski@saic.com>

**References**

Dettling and Buhlmann (2002), *Boosting for Tumor Classification of Gene Expression Data*.

**Examples**

```
# basic interface
r.lb <- LogitBoost(Species ~ ., data=iris, nIter=20)
pred <- predict(r.lb)
prob <- predict(r.lb, type="prob")
d.res <- data.frame(pred, prob)
d.res[1:10, ]

# accuracy increases with nIter (at least for train set)
table(predict(r.lb, iris, type="class", nIter= 2), iris$Species)
table(predict(r.lb, iris, type="class", nIter=10), iris$Species)
table(predict(r.lb, iris, type="class"),          iris$Species)

# example of splitting the data into train and test set
d.set <- SplitTrainTest(iris)
r.lb <- LogitBoost(Species ~ ., data=d.set$train, nIter=10)
table(predict(r.lb, d.set$test, type="class", nIter=2), d.set$test$Species)
table(predict(r.lb, d.set$test, type="class"),          d.set$test$Species)
```

---

Node *Nodes and Splits in an rpart Tree*


---

**Description**

The rpart result object has a complex and compact design. This can make practical use tedious for occasional users as it is difficult to figure out how to access some specific information. The function `Node()` is designed as accessor to the most important properties of a node, being a 'split' or a 'leaf' (aka. 'endnode'). It also serves as base for further convenience functions as e.g. `LeafRates()`.

**Usage**

```
Node(x, node = NULL, type = c("all", "split", "leaf"), digits = 3)
```

**Arguments**

<code>x</code>	fitted model object of class rpart.
<code>node</code>	integer vector, defining the nodes whose details are required.
<code>type</code>	one out of "all" (default), "split", "leaf", where the latter two restrict the result set to splits or end nodes only. Can be abbreviated.
<code>digits</code>	the number of digits for numeric values

**Details**

`Node()` returns detailed information for a single node in the tree. It reports all the data in the summary of a node, but with the option to provide a nodelist. The structure of the result is organised as a list.

**Value**

A list containing:

<code>id</code>	int, id of the node
<code>vname</code>	character, one out of 'leaf' or 'split'
<code>isleaf</code>	logical, TRUE for leaves FALSE else
<code>nobs</code>	integer, number of observation in the node
<code>group</code>	character, the predicted class for the node
<code>ycount</code>	numeric, the number of observation per class in the node
<code>yprob</code>	numeric, the relative frequencies for the each class
<code>nodeprob</code>	the global probability for an observation to fall in the node
<code>complexity</code>	numeric, the complexity parameter for the node
<code>tprint</code>	character, the text to be printed

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[LeafRates](#), [Rules](#)

**Examples**

```
r.rpart <- FitMod(Species ~ ., data=iris, fitfn="rpart")
# return Node nr. 3
Node(r.rpart, node=3)

r.rp <- FitMod(Type ~ ., data = d.glass, fitfn="rpart")
# return all the splits
Node(r.rpart, type="split")
```

---

Over-/Undersample      *Oversample and Undersample*

---

**Description**

For classification purposes we might want to have balanced datasets. If the response variable has not a prevalence of 50%, we can sample records for getting as much response A cases as response B. This is called oversample. Undersample means to sample the (lower) number of cases A from the records of case B.

**Usage**

```
OverSample(x, vname)
UnderSample(x, vname)
```

**Arguments**

x	a data frame containing predictors and response
vname	the name of the response variable to be used to over/undersample

**Value**

a data frame with balanced response variable

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[BestCut](#)

**Examples**

```
OverSample(d.pima2, "diabetes")
```

```
UnderSample(d.pima2, "diabetes")
```

---

 PlotLift

---

*Lift Charts to Compare Binary Predictive Models*


---

**Description**

Provides either a total cumulative response or incremental response rate lift chart for the purposes of comparing the predictive capability of different binary predictive models.

**Usage**

```
PlotLift(modellist, data, targLevel, trueResp, type = "cumulative", sub = "")
```

**Arguments**

modellist	A character vector containing the names of the different models to be compared. The selected models must have the same y variable that must be a binary factor, and have been estimated using the same data set.
data	The dataframe that constitutes the comparison sample. If this dataframe is not the same as the dataframe used to estimate models, the dataframe must contain all the variables used in the models to be compared.
targLevel	The label for the level of the binary factor of interest. For example, in a database marketing application, this level could be "Yes" for a variable that takes on the values "Yes" and "No" to indicate if a customer responded favorably to a promotion offer.
trueResp	The true rate of the target level for the master database the estimation and comparison dataframes were originally drawn from.
type	A character string that must either have the value of "cumulative" (to produce a total cumulative response chart) or "incremental" (to produce an incremental response rate chart).
sub	A sub-title for the plot, typically to identify the sample used.

**Details**

Lift charts are a commonly used tool in business data mining applications. They are used to assess how well a model is able to predict a desirable (from an organization's point-of-view) response on the part of a customer compared to alternative estimated models and a benchmark model of approaching customers randomly. The total cumulative response chart shows the percentage of the total response the organization would receive from only contacting a given percentage (grouped by deciles) of its entire customer base. This chart is best for selecting between alternative models, and in predicting the revenues the organization will receive by contacting a given percentage of their customers that the model predicts are most likely to favorably respond. The incremental response rate chart provides the response rate among each of ten decile groups of the organization's customers, with the decile groups ordered by their estimated likelihood of a favorable response.

**Value**

The function returns the sample response invisibly.

**Author(s)**

original Dan Putler, tweaks Andri Signorell <andri@signorell.net>

**Examples**

```
d.pim <- SplitTrainTest(d.pima, p = 0.2)

r.rp <- FitMod(diabetes ~ pregnant + glucose + pressure + triceps
              + insulin + mass + pedigree + age
              , data=d.pim$train, fitfn="rpart")

r.glm <- FitMod(diabetes ~ pregnant + glucose + pressure + triceps
               + insulin + mass + pedigree + age
               , data=d.pim$train, fitfn="logit")

r.nn <- FitMod(diabetes ~ pregnant + glucose + pressure + triceps
              + insulin + mass + pedigree + age
              , data=d.pim$train, fitfn="nnet")

oldpar <- par(mfrow=c(1,2))
PlotLift(c("r.rp", "r.glm", "r.nn"), data = d.pim$train,
         targLevel = "pos", trueResp =0.34, type = "cumulative")
PlotLift(c("r.rp", "r.glm", "r.nn"), data = d.pim$train,
         targLevel = "pos", trueResp =0.34, type = "incremental")
par(oldpar)
```

---

predict.zeroinfl

*Methods for zeroinfl Objects*

---

**Description**

Methods for extracting information from fitted zero-inflated regression model objects of class "zeroinfl".

**Usage**

```
## S3 method for class 'zeroinfl'
predict(object, newdata,
        type = c("response", "prob", "count", "zero"), na.action = na.pass,
        at = NULL, ...)
## S3 method for class 'zeroinfl'
residuals(object, type = c("pearson", "response"), ...)

## S3 method for class 'zeroinfl'
coef(object, model = c("full", "count", "zero"), ...)
```



```
## S3 method for class 'zeroinfl'
vcov(object, model = c("full", "count", "zero"), ...)

## S3 method for class 'zeroinfl'
terms(x, model = c("count", "zero"), ...)
## S3 method for class 'zeroinfl'
model.matrix(object, model = c("count", "zero"), ...)
```

## Arguments

object, x	an object of class "zeroinfl" as returned by <a href="#">zeroinfl</a> .
newdata	optionally, a data frame in which to look for variables with which to predict. If omitted, the original observations are used.
type	character specifying the type of predictions or residuals, respectively. For details see below.
na.action	function determining what should be done with missing values in newdata. The default is to predict NA.
at	optionally, if type = "prob", a numeric vector at which the probabilities are evaluated. By default 0:max(y) is used where y is the original observed response.
model	character specifying for which component of the model the terms or model matrix should be extracted.
...	currently not used.

## Details

A set of standard extractor functions for fitted model objects is available for objects of class "zeroinfl", including methods to the generic functions [print](#) and [summary](#) which print the estimated coefficients along with some further information. The [summary](#) in particular supplies partial Wald tests based on the coefficients and the covariance matrix (estimated from the Hessian in the numerical optimization of the log-likelihood). As usual, the [summary](#) method returns an object of class "summary.zeroinfl" containing the relevant summary statistics which can subsequently be printed using the associated [print](#) method.

The methods for [coef](#) and [vcov](#) by default return a single vector of coefficients and their associated covariance matrix, respectively, i.e., all coefficients are concatenated. By setting the [model](#) argument, the estimates for the corresponding model components can be extracted.

Both the [fitted](#) and [predict](#) methods can compute fitted responses. The latter additionally provides the predicted density (i.e., probabilities for the observed counts), the predicted mean from the count component (without zero inflation) and the predicted probability for the zero component. The [residuals](#) method can compute raw residuals (observed - fitted) and Pearson residuals (raw residuals scaled by square root of variance function).

The [terms](#) and [model.matrix](#) extractors can be used to extract the relevant information for either component of the model.

A [logLik](#) method is provided, hence [AIC](#) can be called to compute information criteria.

**Author(s)**

Achim Zeileis <Achim.Zeileis@R-project.org>

**See Also**

[zeroinfl](#)

**Examples**

```
data("bioChemists", package = "ModTools")

fm_zip <- zeroinfl(art ~ ., data = bioChemists)
plot(residuals(fm_zip) ~ fitted(fm_zip))

coef(fm_zip)
coef(fm_zip, model = "count")

summary(fm_zip)
logLik(fm_zip)
```

---

PredictCI

*Confidence Intervals for Predictions of a GLM*

---

**Description**

Provides confidence intervals for predictions of a GLM.

**Usage**

```
PredictCI(mod, newdata, conf.level = 0.95)
```

**Arguments**

mod	the binomial model
newdata	the data to be predicted
conf.level	confidence level of the interval. Default is 0.95.

**Details**

The confidence intervals for predictions are calculated with the se of the model and the normal quantile.

**Value**

a matrix with 3 columns for the fit, the lower confidence interval and the upper confidence interval

**Author(s)**

Andri Signorell <andri@signorell.net>

## References

<https://stackoverflow.com/questions/14423325/confidence-intervals-for-predictions-from-logistic-reg>

## See Also

[FitMod](#)

## Examples

```
r.logit <- FitMod(diabetes ~ age, d.pima, fitfn = "logit")
head(PredictCI(r.logit, newdata=d.pima))
```

---

RefLevel

*Used Reference Levels in a Linear Model*

---

## Description

Returns all the reference levels in the factors used in a linear model. It is customer friendly to report also the reference level in lm summaries, which normally are suppressed.

## Usage

```
RefLevel(x)
```

## Arguments

x                   lm object, linear model with factors as predictors.

## Details

For reporting tables of linear models we might want to include an information about the used reference levels, which remain uncommented in the default lm result output. RefLevel() allows to add a footnote or integrate the reference levels in the coefficient table.

## Value

a named vector containing the reference levels of all factors

## Note

It's not clear how general the used algorithm is for more exotic models. [dummy.coef](#) could in such cases be an alternative.

## Author(s)

Andri Signorell <[andri@signorell.net](mailto:andri@signorell.net)>

**See Also**

`dummy.coef`, [Response](#), [relevel](#), [lm](#)

**Examples**

```
RefLevel(lm(breaks ~ wool + tension, data = warpbreaks))
```

---

Response

*Extract the Response from Several Models*

---

**Description**

Time after time, in the course of our daily work, we experience that the response variable is hidden very deeply in the object. This again leads to superfluous consultation of the documentation. `Response()` relieves us of this work.

**Usage**

```
Response(x, ...)
```

**Arguments**

<code>x</code>	the model to use
<code>...</code>	more arguments

**Details**

The function implements the extraction of the response variables for all the models listed in the package's help text.

**Value**

the response of model `x`

**Author(s)**

Andri Signorell <[andri@signorell.net](mailto:andri@signorell.net)>

**See Also**

[model.frame](#), [model.response](#), [RefLevel](#)

**Examples**

```
r.rpart <- FitMod(diabetes ~ ., d.pima, fitfn="rpart")
Response(r.rpart)

# up to the attribute "response" this is the same
identical(StripAttr(Response(r.rpart), "response"),
          model.response(model.frame(r.rpart)))
```

**Description**

For poisson models with mild violation of the distribution assumption that the variance equals the mean, Cameron and Trivedi (2009) recommended using robust standard errors for the parameter estimates. The function uses the function `vcovHC` from the package **sandwich** to obtain the robust standard errors and calculate the p-values accordingly. It returns a matrix containing the usual results in the model summary, comprising the parameter estimates, their robust standard errors, p-values, extended with the 95% confidence interval.

**Usage**

```
RobSummary(mod, conf.level = 0.95, type = "HC0")
```

**Arguments**

<code>mod</code>	the model for which robust standard errors should be calculated
<code>conf.level</code>	the confidence level, default is 95%.
<code>type</code>	a character string specifying the estimation type. Details in <code>vcovHC()</code> .

**Details**

Further details in <https://stats.oarc.ucla.edu/r/dae/poisson-regression/>

**Value**

a  $p \times 6$  matrix with columns for the estimated coefficient, its standard error, t- or z-statistic, the corresponding (two-sided) p-value, the lower and upper confidence interval.

**Author(s)**

Andri Signorell <[andri@signorell.net](mailto:andri@signorell.net)>

**References**

Cameron, A. C. and Trivedi, P. K. (2009) *Microeconometrics Using Stata*. College Station, TX: Stata Press.

**See Also**

[summary.lm](#), [summary.glm](#)

**Examples**

```
r.lm <- lm(Fertility ~ ., swiss)
RobSummary(r.lm)
```

---

ROC

*Build a ROC curve*

---

## Description

This is a wrapper to the main function `pROC` of the **pROC** package (by Xavier Robin et al.). It builds a ROC curve and returns a "roc" object, a list of class "roc".

## Usage

```
ROC(x, resp = NULL, ...)
```

## Arguments

<code>x</code>	a model object, or the predicted probabilities, when <code>resp</code> is not <code>NULL</code> .
<code>resp</code>	the response
<code>...</code>	all arguments are passed to <code>roc()</code> .

## Details

Partial ROC is calculated following Peterson et al. (2008; [doi:10.1016/j.ecolmodel.2007.11.008](https://doi.org/10.1016/j.ecolmodel.2007.11.008)). This function is a modification of the `PartialROC` function, available at <https://github.com/narayanibarve/ENMGadgets>.

## Value

A data.frame containing the AUC values and AUC ratios calculated for each iteration.

## Author(s)

Andri Signorell <[andri@signorell.net](mailto:andri@signorell.net)>

## References

Peterson, A.T. et al. (2008) Rethinking receiver operating characteristic analysis applications in ecological niche modeling. *Ecol. Modell.*, 213, 63-72.

## See Also

[pROC](#)

**Examples**

```

r.glm <- FitMod(diabetes ~ ., data = d.pima, fitfn="logit")
ROC(r.glm)

# plot ROC curves for a list of models
r.rp <- FitMod(diabetes ~ ., data = d.pima, fitfn="rpart")

# combine models to a list
mlst <- list(r.glm, r.rp)

# do the plot
for(i in seq_along(mlst))
  if(i==1){
    plot(ROC(mlst[[i]], grid=TRUE, col=c(hred, hblue)[i]))
  } else {
    lines(ROC(mlst[[i]], col=c(hred, hblue)[i]))
  }

```

Rules

*Extract Rules from 'rpart' Object***Description**

Extract rules from an rpart object. This can be useful, if the rules must be implemented in another system. The rules contain all the criteria for the binary splits of an rpart tree from the root node down to the specified leaf.

**Usage**

```
Rules(x, node = NULL, leafonly = FALSE)
```

**Arguments**

x	the rpart object to extract the rules from
node	integer vector, defining the nodes whose details are required.
leafonly	boolean, defining if only the rules leading to end nodes ("leafs") should be returned.

**Details**

The function builds upon the original function [path.rpart](#), which is bulky in some situations.

**Value**

a list with the rules

frame	the frame of the rpart
ylevels	the y values of the node
ds.size	the size of the dataset
path	a list of character vecotrs containing the rules

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[rpart](#), [path.rpart](#)

**Examples**

```
r.rp <- FitMod(diabetes ~ ., data=d.pima, fitfn="rpart")
Rules(r.rp)
```

---

SplitTrainTest

*Split DataFrame in Train and Test Sample*

---

**Description**

For modeling we usually split our data frame in a train sample, where we train our model on, and a test sample, where we test, how good it works. This function splits a given data frame in two parts, one being the training sample and the other the test sample in form of a list with two elements.

**Usage**

```
SplitTrainTest(x, p = 0.1, seed = NULL, logical = FALSE)
```

**Arguments**

x	data.frame
p	proportion for test sample. Default is 10%.
seed	initialization for random number generator.
logical	logical, defining if a logical vector should be returned or the list with train and test data. Default is FALSE.

**Details**

In order to obtain reasonable models, we should ensure two points. The dataset must be large enough to yield statistically meaningful results and it should be representative of the data set as a whole. Assuming that our test set meets the preceding two conditions, our goal is to create a model that generalizes well to new data. We are aiming for a model that equally well predicts training and test data. We should never train on test data. If we are seeing surprisingly good results on the evaluation metrics, it might be a sign that we're accidentally training on the test set.

**Value**

If `logical` is `FALSE` a list with two data frames, `train` and `test`, of the same structure as the given data in `x`  
if `logical` is `TRUE` a logical vector containing `nrow` elements of `TRUE` and `FALSE`



**Author(s)**

Andri Signorell <andri@signorell.net>

**Examples**

```
SplitTrainTest(d.pima)
```

---

TModC

*Compare Classification Models*


---

**Description**

For the comparison of several classification models, the AUC values and BrierScore values of the models are determined and tabulated. Both the absolute values and the relative values are reported, each related to the model with the highest corresponding value.

**Usage**

```
TModC(..., newdata = NULL, reference = NULL, ord = NULL)
```

```
## S3 method for class 'TModC'
plot(x, col = NULL, args.legend = NULL,...)
```

**Arguments**

...	the models to be compared
x	TModC object to plot
newdata	the data to use for predicting. If not provided, the <code>model.frame</code> will be used.
reference	the reference values
ord	character defining the order of the results table, can be any of "auc", "bs", "auc_p", "bs_p", "bs_rnk", "auc_rnk", "ensemble" (using the mean of "auc_p" and "bs_p" for the ranking).
col	the color for the lines in the ROC plot
args.legend	the legend to be placed in the ROC plot

**Value**

a matrix with the columns

auc	absolute value of area under the ROC curve (AUC)
auc_p	percentage of the auc based on the best observed AUC
auc_rnk	the rank of the auc
bs	absolute value of the Brier score
bs_p	percentage of the Brier score based on the best observed BS
bs_rnk	the rank of the BS
auc_grnk	character representation of the AUC rank
bs_grnk	character representation of the BS rank

**Author(s)**

Andri Signorell <andri@signorell.net>

**See Also**

[TMod](#), [BrierScore](#), [AUC](#), [ROC](#)

**Examples**

```
d.pim <- SplitTrainTest(d.pima, p = 0.2)
mdiab <- formula(diabetes ~ pregnant + glucose + pressure + triceps +
                insulin + mass + pedigree + age)

r.glm <- FitMod(mdiab, data=d.pim$train, fitfn="logit")
r.rp <- FitMod(mdiab, data=d.pim$train, fitfn="rpart")
mods <- list(glm=r.glm, rp=r.rp)

# the table with the measures
(tm <- TModC(mods, ord="auc"))

# plotting the ROC curves
plot(tm, col=c("darkmagenta", "dodgerblue"))
```

---

Tobit

*Tobit Regression*

---

**Description**

Fitting and testing Tobit regression models for censored data.

**Usage**

```
Tobit(formula, left = 0, right = Inf, dist = "gaussian",
       subset = NULL, data = list(), ...)
```

**Arguments**

formula	a symbolic description of a regression model of type $y \sim x_1 + x_2 + \dots$
left	left limit for the censored dependent variable $y$ . If set to $-\text{Inf}$ , $y$ is assumed not to be left-censored.
right	right limit for the censored dependent variable $y$ . If set to $\text{Inf}$ , the default, $y$ is assumed not to be right-censored.
dist	assumed distribution for the dependent variable $y$ . This is passed to <a href="#">survreg</a> , see the respective man page for more details.
subset	a specification of the rows to be used.
data	a data frame containing the variables in the model.
...	further arguments passed to <a href="#">survreg</a> .

**Details**

The function `Tobit` is an alias for the **AER** function `tobit` (Achim Zeileis <Achim.Zeileis@R-project.org>). All details can be found there.

**Value**

An object of class "Tobit" inheriting from class "survreg".

**Author(s)**

Andri Signorell

**Examples**

```
# still to do
```

---

Tune

*Tune Classifiers*

---

**Description**

Some classifiers benefit more from adjusted parameters to a particular dataset than others. However, it is often not clear from the beginning how the parameters have to be determined. What often only remains is a grid search when several parameters have to be found in combination. The present function uses a grid search approach for the decisive arguments (typically for a neural network, a random forest or a classification tree). However it's not restricted to these models, any model fulfilling weak interface standards could be provided.

**Usage**

```
Tune(x, ..., testset = NULL, keepmod = TRUE)
```

**Arguments**

<code>x</code>	the model to be tuned, best (but not necessarily) trained with <code>FitMod</code> .
<code>...</code>	a list of parameters, containing the values to be used for a grid search.
<code>testset</code>	a testset containing all variables required in the model to be used for calculating independently the accuracy (normally a subset of the original dataset).
<code>keepmod</code>	logical, defining if all fitted models should be returned in the result set. Default is TRUE. (Keep an eye on your RAM!)

## Details

The function creates a n-dimensional grid according to the given parameters and calculates the model with the combinations of all the parameters. The accuracy for the models are calculated insample and on a test set, if one has been provided.

It makes sense to avoid overfitting to provide a test set to also be evaluated. A matrix with all combination of the values for the given parameters, sorted by accuracy, either by the accuracy achieved in the test set or the insample accuracy is returned.

## Value

a matrix with all supplied parameters and a column "acc" and "test\_acc" (if a test set has been provided)

## Author(s)

Andri Signorell <andri@signorell.net>

## Examples

```
d.pim <- SplitTrainTest(d.pima, p = 0.2)
mdiab <- formula(diabetes ~ pregnant + glucose + pressure + triceps
                + insulin + mass + pedigree + age)

# tune a neural network for size and decay
r.nn <- FitMod(mdiab, data=d.pim$train, fitfn="nnet")
(tu <- Tune(r.nn, size=12:17, decay = 10^(-4:-1), testset=d.pim$test))

# tune a random forest
r.rf <- FitMod(mdiab, data=d.pim$train, fitfn="randomForest")
(tu <- Tune(r.rf, mtry=seq(2, 20, 2), testset=d.pim$test))

# tune a SVM model
r.svm <- FitMod(mdiab, data=d.pim$train, fitfn="svm")

tu <- Tune(r.svm,
          kernel=c("radial", "sigmoid"),
          cost=c(0.1,1,10,100,1000),
          gamma=c(0.5,1,2,3,4), testset=d.pim$test)

# let's get some more quality measures
tu$modpar$Sens <- sapply(tu$mods, Sens)      # Sensitivity
tu$modpar$Spec <- sapply(tu$mods, Spec)     # Specificity
Sort(tu$modpar, ord="test_acc", decreasing=TRUE)
```

**Description**

Variable importance is an expression of the desire to know how important a variable is within a group of predictors for a particular model. But in general it is not a well defined concept, say there is no theoretically defined variable importance metric. Nevertheless, there are some approaches that have been established in practice for some regression and classification algorithms. The present function provides an interface for calculating variable importance for some of the models produced by `FitMod`, comprising linear models, classification trees, random forests, C5 trees and neural networks. The intention here is to provide reasonably homogeneous output and plot routines.

**Usage**

```
VarImp(x, scale = FALSE, sort = TRUE, ...)

## S3 method for class 'FitMod'
VarImp(x, scale = FALSE, sort = TRUE, type=NULL, ...)
## Default S3 method:
VarImp(x, scale = FALSE, sort = TRUE, ...)

## S3 method for class 'VarImp'
plot(x, sort = TRUE, maxrows = NULL,
      main = "Variable importance", ...)

## S3 method for class 'VarImp'
print(x, digits = 3, ...)
```

**Arguments**

<code>x</code>	the fitted model
<code>scale</code>	logical, should the importance values be scaled to 0 and 100?
<code>...</code>	parameters to pass to the specific <code>VarImp</code> methods
<code>sort</code>	the name of the column, the importance table should be ordered after
<code>maxrows</code>	the maximum number of rows to be reported
<code>main</code>	the main title for the plot
<code>type</code>	some models have more than one type available to produce a variable importance. Linear models accept one of "lmg", "pmvd", "first", "last", "betasq", "pratt".
<code>digits</code>	the number of digits for printing the "VarImp" table

**Details**

**Linear Models:** For linear models there's a fine package **relaimpo** available on CRAN containing several interesting approaches for quantifying the variable importance. See the original documentation.

**rpart, Random Forest:** `VarImp.rpart` and `VarImp.randomForest` are wrappers around the importance functions from the **rpart** or **randomForest** packages, respectively.

**C5.0:** C5.0 measures predictor importance by determining the percentage of training set samples that fall into all the terminal nodes after the split. For example, the predictor in the first split automatically has an importance measurement of 100 percent since all samples are affected by this split. Other predictors may be used frequently in splits, but if the terminal nodes cover only a handful of training set samples, the importance scores may be close to zero. The same strategy is applied to rule-based models and boosted versions of the model. The underlying function can also return the number of times each predictor was involved in a split by using the option `metric="usage"`.

**Neural Networks:** The method used here is "Garson weights".

**SVM, GLM, Multinom:** There are no implementations for these models so far.

**Value**

A data frame with class `c("VarImp.train", "data.frame")` for `VarImp.train` or a matrix for other models.

**Author(s)**

Andri Signorell <andri@signorell.net>

**References**

Quinlan, J. (1992). Learning with continuous classes. Proceedings of the 5th Australian Joint Conference On Artificial Intelligence, 343-348.

---

zeroinfl

*Zero-inflated Count Data Regression*

---

**Description**

Fit zero-inflated regression models for count data via maximum likelihood.

**Usage**

```
zeroinfl(formula, data, subset, na.action, weights, offset,
  dist = c("poisson", "negbin", "geometric"),
  link = c("logit", "probit", "cloglog", "cauchit", "log"),
  control = zeroinfl.control(...),
  model = TRUE, y = TRUE, x = FALSE, ...)
```

## Arguments

formula	symbolic description of the model, see details.
data, subset, na.action	arguments controlling formula processing via <a href="#">model.frame</a> .
weights	optional numeric vector of weights.
offset	optional numeric vector with an a priori known component to be included in the linear predictor of the count model. See below for more information on offsets.
dist	character specification of count model family (a log link is always used).
link	character specification of link function in the binary zero-inflation model (a binomial family is always used).
control	a list of control arguments specified via <a href="#">zeroinfl.control</a> .
model, y, x	logicals. If TRUE the corresponding components of the fit (model frame, response, model matrix) are returned.
...	arguments passed to <a href="#">zeroinfl.control</a> in the default setup.

## Details

Zero-inflated count models are two-component mixture models combining a point mass at zero with a proper count distribution. Thus, there are two sources of zeros: zeros may come from both the point mass and from the count component. Usually the count model is a Poisson or negative binomial regression (with log link). The geometric distribution is a special case of the negative binomial with size parameter equal to 1. For modeling the unobserved state (zero vs. count), a binary model is used that captures the probability of zero inflation. In the simplest case only with an intercept but potentially containing regressors. For this zero-inflation model, a binomial model with different links can be used, typically logit or probit.

The formula can be used to specify both components of the model: If a formula of type  $y \sim x_1 + x_2$  is supplied, then the same regressors are employed in both components. This is equivalent to  $y \sim x_1 + x_2 \mid x_1 + x_2$ . Of course, a different set of regressors could be specified for the count and zero-inflation component, e.g.,  $y \sim x_1 + x_2 \mid z_1 + z_2 + z_3$  giving the count data model  $y \sim x_1 + x_2$  conditional on (1) the zero-inflation model  $y \sim z_1 + z_2 + z_3$ . A simple inflation model where all zero counts have the same probability of belonging to the zero component can be specified by the formula  $y \sim x_1 + x_2 \mid 1$ .

Offsets can be specified in both components of the model pertaining to count and zero-inflation model:  $y \sim x_1 + \text{offset}(x_2) \mid z_1 + z_2 + \text{offset}(z_3)$ , where  $x_2$  is used as an offset (i.e., with coefficient fixed to 1) in the count component and  $z_3$  analogously in the zero-inflation component. By the rule stated above  $y \sim x_1 + \text{offset}(x_2)$  is expanded to  $y \sim x_1 + \text{offset}(x_2) \mid x_1 + \text{offset}(x_2)$ . Instead of using the `offset()` wrapper within the formula, the `offset` argument can also be employed which sets an offset only for the count model. Thus, `formula = y ~ x1` and `offset = x2` is equivalent to `formula = y ~ x1 + offset(x2) | x1`.

All parameters are estimated by maximum likelihood using `optim`, with control options set in [zeroinfl.control](#). Starting values can be supplied, estimated by the EM (expectation maximization) algorithm, or by `glm.fit` (the default). Standard errors are derived numerically using the Hessian matrix returned by `optim`. See [zeroinfl.control](#) for details.

The returned fitted model object is of class "zeroinfl" and is similar to fitted "glm" objects. For elements such as "coefficients" or "terms" a list is returned with elements for the zero and count component, respectively. For details see below.

A set of standard extractor functions for fitted model objects is available for objects of class "zeroinfl", including methods to the generic functions `print`, `summary`, `coef`, `vcov`, `logLik`, `residuals`, `predict`, `fitted`, `terms`, `model.matrix`. See `predict.zeroinfl` for more details on all methods.

## Value

An object of class "zeroinfl", i.e., a list with components including

<code>coefficients</code>	a list with elements "count" and "zero" containing the coefficients from the respective models,
<code>residuals</code>	a vector of raw residuals (observed - fitted),
<code>fitted.values</code>	a vector of fitted means,
<code>optim</code>	a list with the output from the <code>optim</code> call for minimizing the negative log-likelihood,
<code>control</code>	the control arguments passed to the <code>optim</code> call,
<code>start</code>	the starting values for the parameters passed to the <code>optim</code> call,
<code>weights</code>	the case weights used,
<code>offset</code>	a list with elements "count" and "zero" containing the offset vectors (if any) from the respective models,
<code>n</code>	number of observations (with weights > 0),
<code>df.null</code>	residual degrees of freedom for the null model (= n - 2),
<code>df.residual</code>	residual degrees of freedom for fitted model,
<code>terms</code>	a list with elements "count", "zero" and "full" containing the terms objects for the respective models,
<code>theta</code>	estimate of the additional $\theta$ parameter of the negative binomial model (if a negative binomial regression is used),
<code>SE.logtheta</code>	standard error for $\log(\theta)$ ,
<code>loglik</code>	log-likelihood of the fitted model,
<code>vcov</code>	covariance matrix of all coefficients in the model (derived from the Hessian of the <code>optim</code> output),
<code>dist</code>	character string describing the count distribution used,
<code>link</code>	character string describing the link of the zero-inflation model,
<code>linkinv</code>	the inverse link function corresponding to <code>link</code> ,
<code>converged</code>	logical indicating successful convergence of <code>optim</code> ,
<code>call</code>	the original function call,
<code>formula</code>	the original formula,
<code>levels</code>	levels of the categorical regressors,



contrasts	a list with elements "count" and "zero" containing the contrasts corresponding to levels from the respective models,
model	the full model frame (if model = TRUE),
y	the response count vector (if y = TRUE),
x	a list with elements "count" and "zero" containing the model matrices from the respective models (if x = TRUE),

### Author(s)

Achim Zeileis <Achim.Zeileis@R-project.org>

### References

- Cameron, A. Colin and Pravin K. Trivedi. 1998. *Regression Analysis of Count Data*. New York: Cambridge University Press.
- Cameron, A. Colin and Pravin K. Trivedi. 2005. *Microeconometrics: Methods and Applications*. Cambridge: Cambridge University Press.
- Lambert, Diane. 1992. "Zero-Inflated Poisson Regression, with an Application to Defects in Manufacturing." *Technometrics*. **34**(1):1-14
- Zeileis, Achim, Christian Kleiber and Simon Jackman 2008. "Regression Models for Count Data in R." *Journal of Statistical Software*, **27**(8). URL <https://www.jstatsoft.org/v27/i08/>.

### See Also

[zeroinfl.control](#), [glm](#), [glm.fit](#), [glm.nb](#), [hurdle](#)

### Examples

```
## data
data("bioChemists", package = "ModTools")

## without inflation
## ("art ~ ." is "art ~ fem + mar + kid5 + phd + ment")
fm_pois <- glm(art ~ ., data = bioChemists, family = poisson)
fm_qpois <- glm(art ~ ., data = bioChemists, family = quasipoisson)
fm_nb <- MASS::glm.nb(art ~ ., data = bioChemists)

## with simple inflation (no regressors for zero component)
fm_zip <- zeroinfl(art ~ . | 1, data = bioChemists)
fm_zinb <- zeroinfl(art ~ . | 1, data = bioChemists, dist = "negbin")

## inflation with regressors
## ("art ~ . | ." is "art ~ fem + mar + kid5 + phd + ment | fem + mar + kid5 + phd + ment")
fm_zip2 <- zeroinfl(art ~ . | ., data = bioChemists)
fm_zinb2 <- zeroinfl(art ~ . | ., data = bioChemists, dist = "negbin")
```

---

zeroinfl.control      *Control Parameters for Zero-inflated Count Data Regression*

---

### Description

Various parameters that control fitting of zero-inflated regression models using [zeroinfl](#).

### Usage

```
zeroinfl.control(method = "BFGS", maxit = 10000, trace = FALSE,  
  EM = FALSE, start = NULL, ...)
```

### Arguments

method	characters string specifying the method argument passed to <a href="#">optim</a> .
maxit	integer specifying the maxit argument (maximal number of iterations) passed to <a href="#">optim</a> .
trace	logical or integer controlling whether tracing information on the progress of the optimization should be produced (passed to <a href="#">optim</a> ).
EM	logical. Should starting values be estimated by the EM (expectation maximization) algorithm? See details.
start	an optional list with elements "count" and "zero" (and potentially "theta") containing the coefficients for the corresponding component.
...	arguments passed to <a href="#">optim</a> .

### Details

All parameters in [zeroinfl](#) are estimated by maximum likelihood using [optim](#) with control options set in [zeroinfl.control](#). Most arguments are passed on directly to [optim](#), only `trace` is also used within [zeroinfl](#) and `EM/start` control the choice of starting values for calling [optim](#).

Starting values can be supplied, estimated by the EM (expectation maximization) algorithm, or by [glm.fit](#) (the default). Standard errors are derived numerically using the Hessian matrix returned by [optim](#). To supply starting values, `start` should be a list with elements "count" and "zero" and potentially "theta" (for negative binomial components only) containing the starting values for the coefficients of the corresponding component of the model.

### Value

A list with the arguments specified.

### Author(s)

Achim Zeileis <Achim.Zeileis@R-project.org>

### See Also

[zeroinfl](#)

**Examples**

```
## Not run:
data("bioChemists", package = "pscl")

## default start values
fm1 <- zeroinfl(art ~ ., data = bioChemists)

## use EM algorithm for start values
fm2 <- zeroinfl(art ~ ., data = bioChemists, EM = TRUE)

## user-supplied start values
fm3 <- zeroinfl(art ~ ., data = bioChemists,
  start = list(count = c(0.7, -0.2, 0.1, -0.2, 0, 0), zero = -1.7))

## End(Not run)
```

# Index

- \* **classif**
    - LogitBoost, 19
  - \* **datasets**
    - bioChemists, 9
    - d.glass, 14
    - d.pima, 15
  - \* **htest**
    - BreuschPaganTest, 10
  - \* **misc**
    - PlotLift, 23
  - \* **models**
    - CP, 12
    - FitMod, 16
    - Response, 28
    - Rules, 31
    - TModC, 33
    - VarImp, 37
  - \* **model**
    - ROC, 30
    - SplitTrainTest, 32
  - \* **package**
    - ModTools-package, 3
  - \* **regression**
    - predict.zeroinfl, 24
    - Tobit, 34
    - zeroinfl, 38
    - zeroinfl.control, 42
  - \* **tree**
    - LeafRates, 17
    - Node, 21
- AIC, 25  
AUC, 34
- BestCut, 4, 8, 22  
bioChemists, 9  
bondyield, 10  
BreuschPaganTest, 4, 10  
BrierScore, 34
- C5.0, 3  
coef, 25, 40  
coef.zeroinfl (predict.zeroinfl), 24  
CoeffDiffCI, 11  
CP, 4, 12  
currencysubstitution, 10
- d.glass, 14  
d.pima, 15  
d.pima2 (d.pima), 15  
drop1.FitMod (FitMod), 16  
dummy.coef, 27
- extractAIC.zeroinfl (predict.zeroinfl), 24
- FitMod, 3, 16, 27, 35  
fitted, 25, 40  
fitted.zeroinfl (predict.zeroinfl), 24  
formula, 16, 19
- GarsonWeights (VarImp), 37  
glm, 3, 41  
glm.fit, 39, 41, 42  
glm.nb, 41  
growthofmoney, 10
- hurdle, 41
- lda, 3  
LeafRates, 4, 17, 21, 22  
linearHypothesis, 12  
lm, 3, 11, 17, 28  
lmrob, 3  
LogitBoost, 4, 19  
logLik, 25, 40  
logLik.zeroinfl (predict.zeroinfl), 24
- model.frame, 28, 39  
model.matrix, 25, 40

- model.matrix.zeroinfl
  - (predict.zeroinfl), 24
- model.response, 28
- ModTools (ModTools-package), 3
- ModTools-package, 3
- moneydemand, 10
- multinom, 3
  
- naive\_bayes, 3
- ncvTest, 11
- nnet, 3
- Node, 4, 18, 21
  
- optim, 39, 42
- Over-/Undersample, 22
- OverSample, 4
- OverSample (Over-/Undersample), 22
  
- path.rpart, 31, 32
- plot.CP (CP), 12
- plot.FitMod (FitMod), 16
- plot.LeafRates (LeafRates), 17
- plot.TModC (TModC), 33
- plot.VarImp (VarImp), 37
- plotcp, 13
- PlotLift, 4, 23
- polr, 3
- predict, 4, 25, 40
- predict.FitMod (FitMod), 16
- predict.zeroinfl, 24, 40
- PredictCI, 26
- predprob.zeroinfl (predict.zeroinfl), 24
- print, 25, 40
- print.CP (CP), 12
- print.FitMod (FitMod), 16
- print.summary.zeroinfl
  - (predict.zeroinfl), 24
- print.VarImp (VarImp), 37
- print.zeroinfl (zeroinfl), 38
- printcp, 13
- pROC, 30
- Purity (LeafRates), 17
  
- qda, 3
  
- randomForest, 3
- RefLevel, 27, 28
- relevel, 28
- residuals, 25, 40
- residuals.zeroinfl (predict.zeroinfl), 24
- Response, 4, 28, 28
- RobSummary, 4, 29
- ROC, 4, 9, 30, 34
- rpart, 3, 17, 19, 32
- Rules, 4, 18, 22, 31
  
- Splits (Node), 21
- SplitTrainTest, 4, 32
- summary, 25, 40
- summary.FitMod (FitMod), 16
- summary.glm, 29
- summary.lm, 29
- summary.zeroinfl (predict.zeroinfl), 24
- survreg, 34
- svm, 3
  
- terms, 25, 40
- terms.zeroinfl (predict.zeroinfl), 24
- TMod, 34
- TModC, 4, 33
- Tobit, 34
- tobit, 35
- Tune, 4, 35
  
- UnderSample (Over-/Undersample), 22
- unemployment, 10
  
- VarImp, 4, 37
- vcov, 25, 40
- vcov.zeroinfl (predict.zeroinfl), 24
- vcovHC, 29
  
- wages, 10
  
- zeroinfl, 25, 26, 38, 42
- zeroinfl.control, 39, 41, 42, 42